# BONE MARROW AND SPLEEN EXAMINATION

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# HISTOLOGY OF BONE MARROW

## Overview

#### **Bone Marrow**

- Bone marrow sample
- Bone marrow overview
- Bone marrow histology in a core biopsy
- Myeloid cells
- Monocytic and dendritic cells
- Erythroid cells
- Megakaryocytes
- Lymphoid histology
- Bone

#### Spleen

An abbreviated version

## Bone Marrow in The Pathology Lab

#### **Usual Sample**

- Bone marrow smear
- Bone marrow clot
- Bone marrow biopsy (decalcified)



Always add information from

- Clinical history
- Labs (specially CBC)
- Flow cytometry
- Iron stain

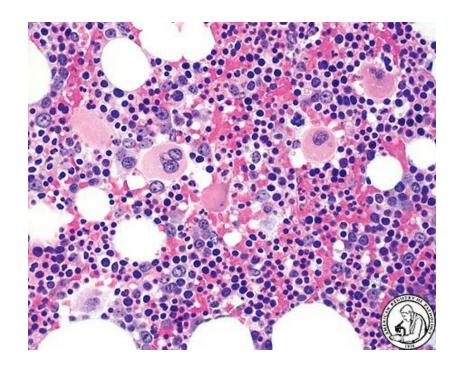
## **Bone Marrow Biopsy**

- The biopsy is used for <u>quantitative</u> evaluation of the marrow
  - Cellularity
  - M:E ratio, it is considered the best estimation (vs differential from smear)
  - Numbers of megakaryocytes
  - Presence of tumors, granulomas, infiltrates
  - Presence of fibrosis, necrosis



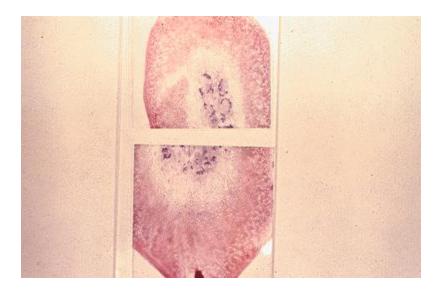
## **Bone Marrow Clot Section**

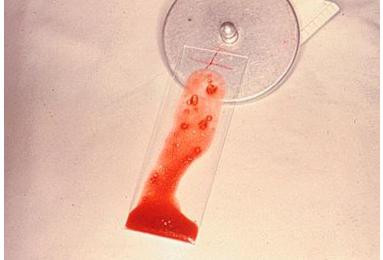
- May also be used for quantitative evaluation, especially if the biopsy is suboptimal
- May give a better evaluation of erythropoiesis, since there is no bone present in the specimen, it can be cut thinner, aiding in the evaluation of red cell precursors
- The best for immunos... if representative



## **Bone Marrow Smears**

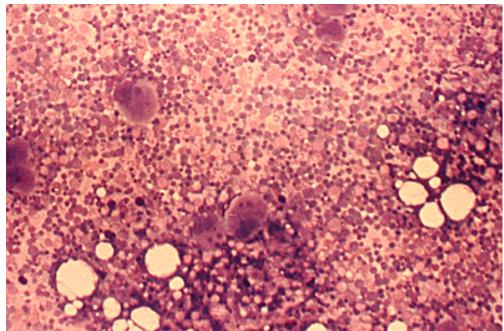
- The smears are used for <u>qualitative</u> evaluation of the marrow
  - cell identification
  - dyspoiesis
  - maturation
  - cytologic abnormalities





## Bone Marrow Smears

- The smears are used for <u>qualitative</u> evaluation of the marrow
  - cell identification
  - dyspoiesis
  - maturation
  - cytologic abnormalities



This bone marrow is from a patient with chronic myelocytic leukemia.

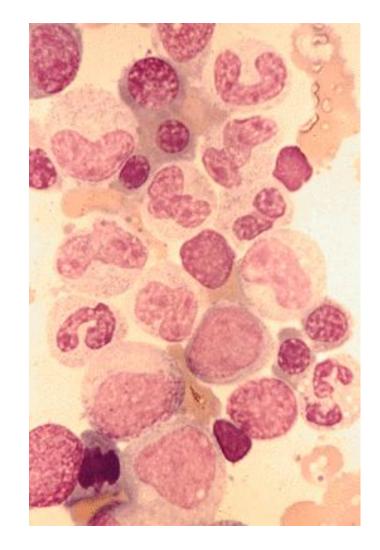
It is hypercellular and contains only a small amount of fat. There are a number of

megakaryocytes, which are the largest cells of the bone marrow. The number of **megakaryocytes** is estimated in the thickest part of the particle and not in the sinusoidal blood. The normal number is 3-10 per low-power field. **More than 15 in one field in the particle is considered increased**. If you have to search for them and find only **1-2 per particle, they are decreased**.

### Bone Marrow Smear – The differential

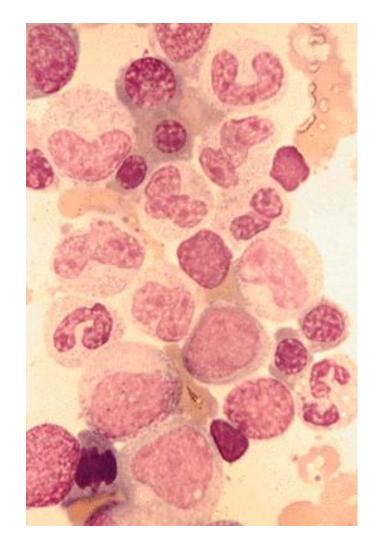
#### **Practical advices**

- Always do your differential with 1,000x magnification.
- · Count at least 200-300 nucleated cells
- Between 40-60 cells can be counted in one field with oil immersion.
- It becomes difficult with 100 or more cells, as we tend to skip or to count the same cells twice.
- Fewer than 10 cells per field indicates sinusoidal blood rather than actual bone marrow particles I divide the field into imaginary quarters and start at the 12-o'clock position and count clockwise.
- Beginners find it easier to count all the granulocytes in the field and then to go back over it and count the lymphocytes, nucleated red cells, plasma cells, and the other types in the same fashion.



### Bone Marrow Smear – The differential

- There are 13 granulocytes:
  - two segs
  - four bands
  - four metamyelocytes
  - two myelocytes
  - one promyelocyte (some promyelocytes do not contain granules)
- Four normoblasts (E5)
- Two late erythroblasts (E4) one of which is in mitosis.
  - There are also three naked nuclei, which are disregarded



## Basic approach in the biopsy

- Cellularity
  - Depending on Pt age
- Myeloid to Erythroid ratio (ME ratio)
  - 1:6 in first week of life
  - 2.5-4:1 (adult physiologic range [3:1])

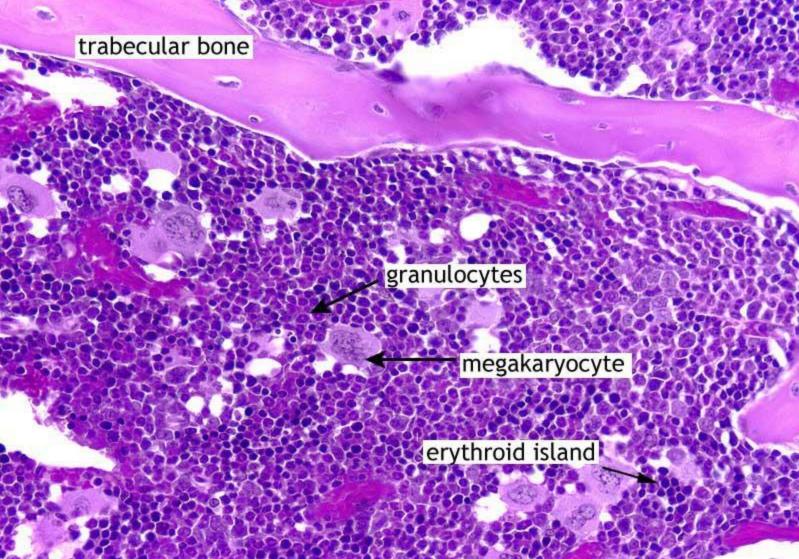


 Increase in either component is reported as myeloid/erythroid "predominant" in the presence of a normal fat:cell ratio, and "hyperplasia" when the cellularity of the bone marrow exceeds 70%.

## How to tell which ones are which?

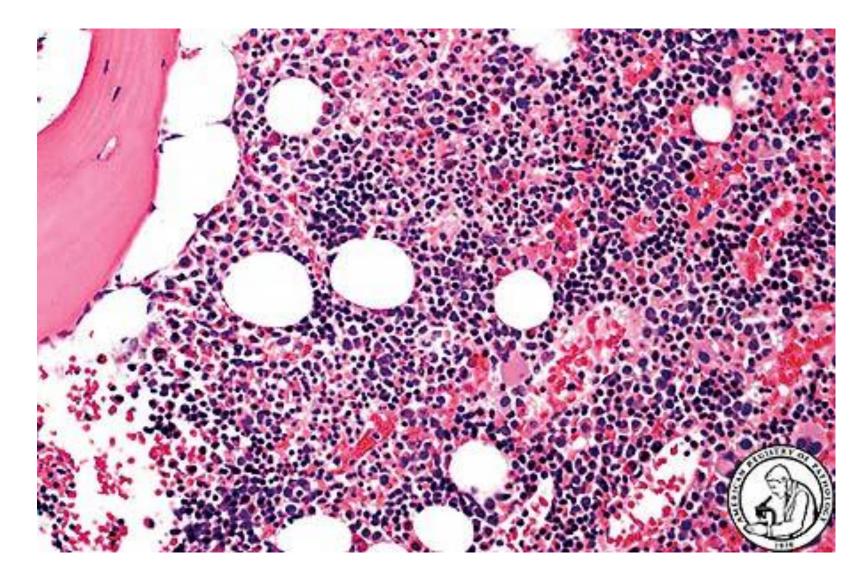
- Erythroid cells are seen as round dark nuclei without much cytoplasm - "black dots" usually with a halo around them
- Myeloid cells have lighter staining nuclei and pink cytoplasm, with maturation you can identify polys and bands (paratrabecular distribution)
- Megakaryocytes are <u>large</u> with multilobated nuclei (perisinusoidal)

## Myeloid vs. Erythroid



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## Erythroid hyperplasia

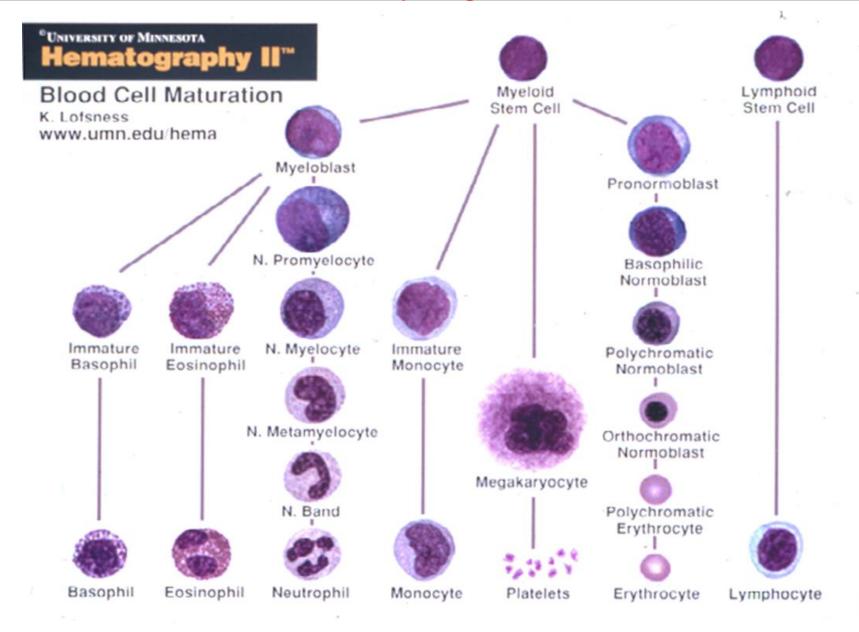


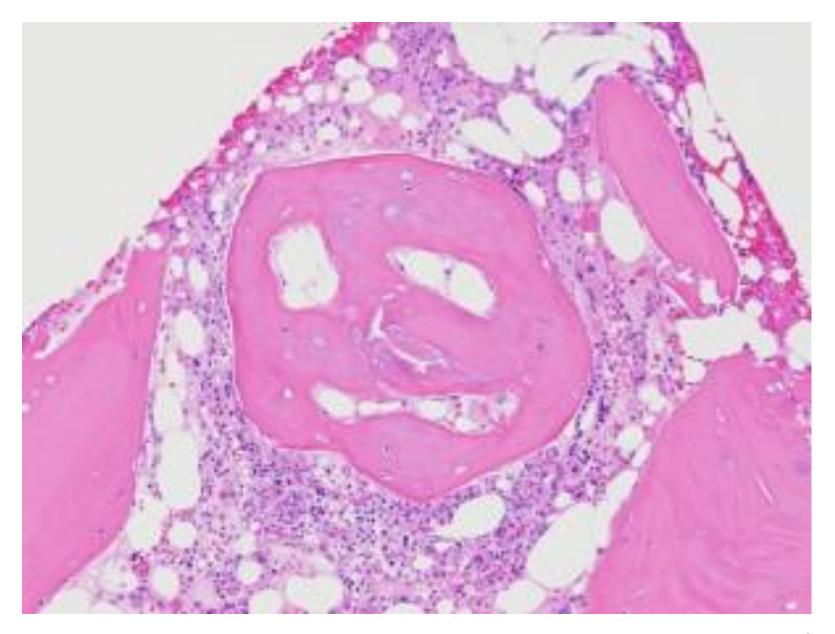
## **Bone marrow lexicon**

- Blast  $\rightarrow$  least differentiated
- Cyte → more differentiated
- Pro  $\rightarrow$  2<sup>nd</sup> cell in maturation sequence
- Meta → 4<sup>th</sup> cell in maturation sequence (if 4 maturation stages)

	Table 4 – Nomenclature								
PREFIX	SUFFIX	MYELOCYTIC SERIES	MONOCYTIC SERIES	MEGAKARYOCYTIC SERIES	ERYTHROCYTIC (RUBRICYTIC) SERIES	LYMPHOCYTIC SERIES	PLASMOCYTIC SERIES		
pro meta	blast cyte cyte cyte	myeloblast promyelocyte myelocyte metamyelocyte	monoblast promonocyte monocyte	megakaryoblast promegakaryocyte megakaryocyte metamegakaryocyte	rubriblast prorubricyte rubricyte metarubricyte	lymphoblast prolymphocyte lymphocyte	plasmoblast proplasmocyte plasmocyte		

#### Bone marrow, differentiation and cytological features





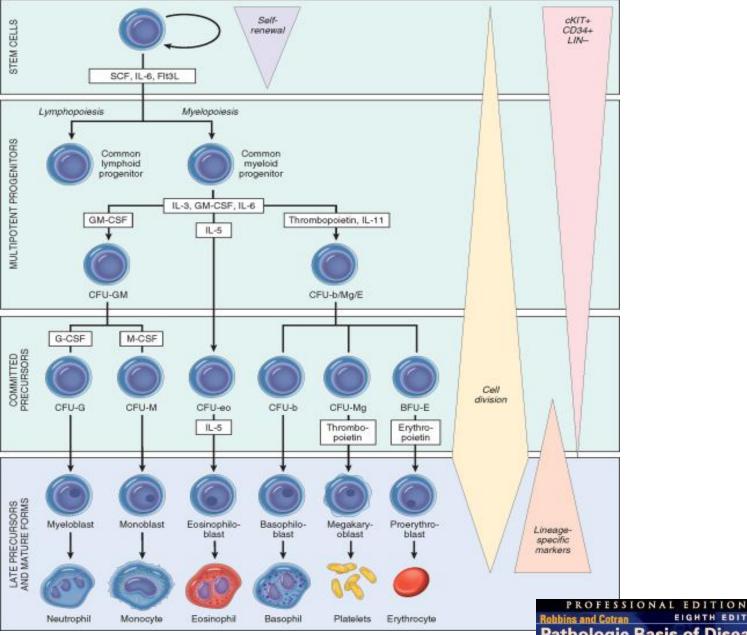
www.pathtalk.org

## Fetal hematopoiesis

- First detected in the yolk sac (2<sup>nd</sup> to 3<sup>rd</sup> week), and exclusively produces nucleated red blood cells
- Then dorsal aorta, liver (6<sup>th</sup> week), spleen and bone marrow (14<sup>th</sup> weeks)
- Hematopoietic stem cells (CD34, c-kit, Thy1) and CD34 receptors
- The mesenchymal component (adipose tissue, bone and fibroblast) comes from primitive mesenchymal stem cells.

### **Microenvironment Controls Differentiation**

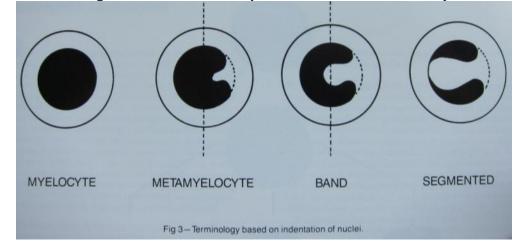
- Immature granulocytic cells have paratrabecular arrangement.
- T-cells and macrophages (IL-6, G-CSF, M-CSF) produce several cytokines that regulate the microenvironment
- Specific cytokines promote lineage specific proliferation:
  - G-CSF → Granulocytes
  - M- CSF → Monocytes/macrocytic lineage
  - IL-5  $\rightarrow$  Eosinophil/basophil production



**Pathologic Basis of Disease** KUMAR . ABBAS . FAUSTO . ASTER

EIGHTH EDITION

 Granulopoietic cycle within the bone marrow takes 10 to 14 days but can be accelerated by cytokines (G/GM-CSF)



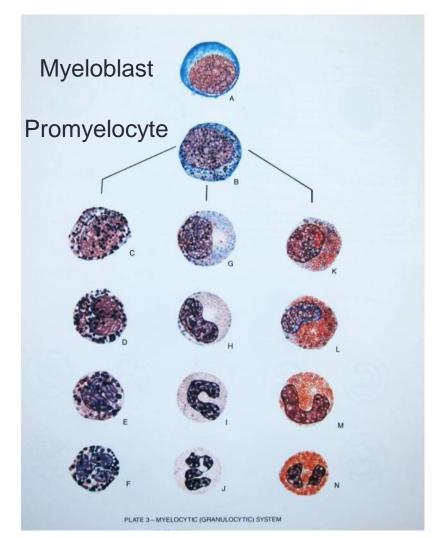
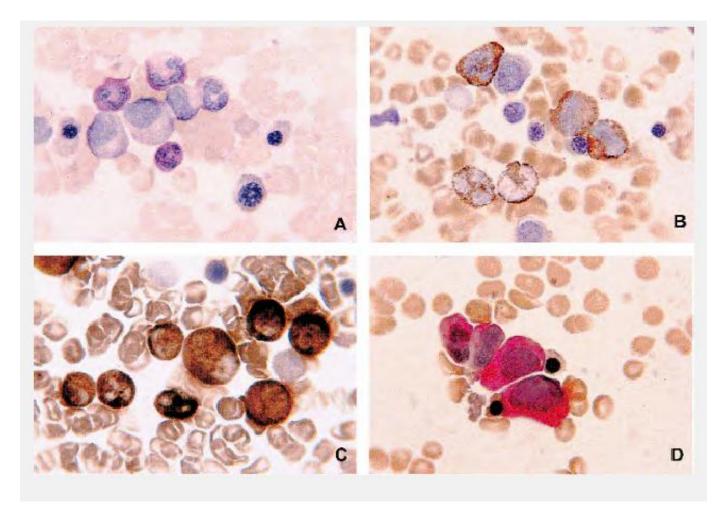


TABLE 2. Types of myeloid elements and their normal range in the bone marrow

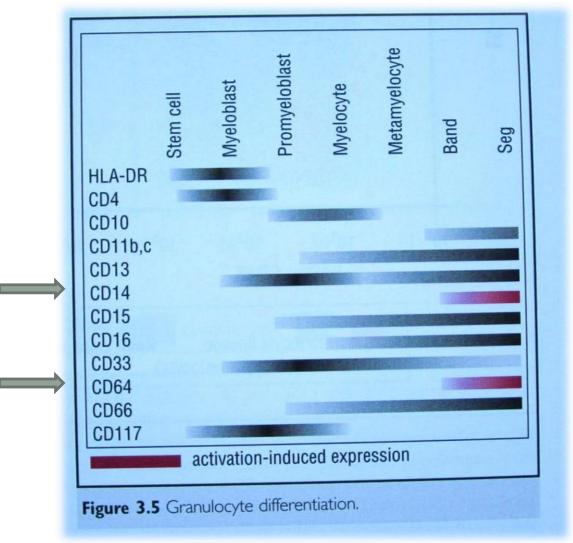
Cell type	Range
Myeloblasts	0% to 2%
Promyelocytes	2% to 5%
Myelocytes (neutrophilic)	9% to 16%
Metamyelocytes	7% to 23%
Band Forms	8% to 15%
Neutrophils	4% to 10%
Myelocytes (eosinophilic)	0% to 2%
Band	0% to 2%
Mature	0% to 2%
Monocytes/macrophages	0% to 3%
Basophils	0% to 1%
Mast Cells	0% to 2%



A. PAS, B. Sudan black, C. Myeloperoxidase, D. Alpha-naphthol AS-D chloroacetate

Table 1-2 GRANULOPOIESIS <sup>a</sup>						
Stage of Maturation	Morphology	Cytochemical/Immunophenotypic Properties				
Myeloblast	High nuclear to cytoplasmic ratio Blastic, dispersed chromatin Agranular, minimally granular cytoplasm	Myeloperoxidase + or – Most myeloblasts are CD34+, HLA-DR+, and coexpress myeloid+ lineage antigen such as CD33				
Promyelocyte	Eccentric nucleus with prominent para- nuclear hof (pale zone) Sparse, concentrated azurophilic granules	Myeloperoxidase + Typically CD34 –, HLA-DR –, and myeloid antigen + (e.g CD33, CD13)				
Neutrophilic myelocyte	Round nucleus with condensed chromatin Moderate to abundant secondary (specific) granules which give the cytoplasm a finely granular pink appearance	Myeloperoxidase +, leukocyte alkaline phosphatase + Myeloid antigen + (CD34 –, HLA-DR –)				
Neutrophilic metamyelo- cyte	Indented nucleus, condensed chromatin Cytoplasm packed with granules with predominance of secondary granules	Myeloperoxidase +, leukocyte alkaline phosphatase + Myeloid antigen + (CD34-, HLA-DR-)				
Band neutro- phil	Horseshoe-shaped mature nucleus lacking discrete indentations Cytoplasm packed with granules with pre- dominance of secondary granules; gelatinous granules also present	Myeloperoxidase +, leukocyte alkaline phosphatase + Myeloid antigen + (CD34 – HLA-DR –)				
Neutrophil	<ul> <li>3-5 discrete nuclear lobes</li> <li>Highly condensed chromatin</li> <li>Cytoplasm packed with granules with pre- dominance of secondary granules; gelatin- ous granules also present</li> </ul>	Myeloperoxidase+, leukocyte alkaline phosphatase + Myeloid antigen + (CD34 –, HLA-DR –)				

<sup>a</sup>Data from references 39, 46, and 62.

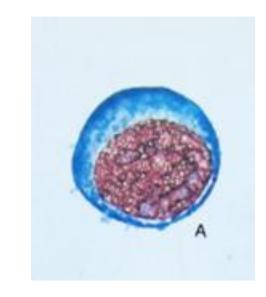


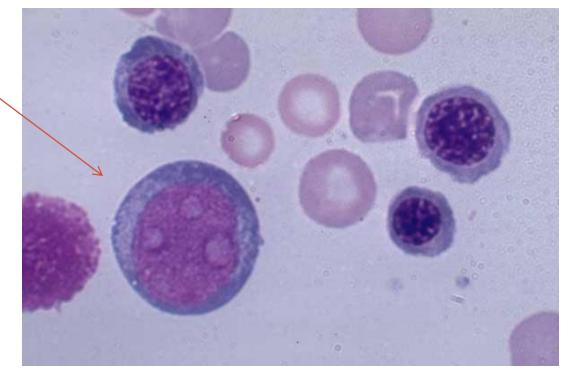
## Neutrophilic Leukocytes

- Maturation is characterized by development of dark-blue (primary) granules that later on is replaced by secondary granules that differ in their size and staining pattern
  - Neutrophil small granules that stain with light blue and pink
    - Also: Gelatinase (tertiary) granule, which lacks both myeloperoxidase and lactoferrin, but contains gelatinase, acetyltransferase, and lysozyme
  - Basophil large basophilic granules
  - Eosinophil

## Myeloblast

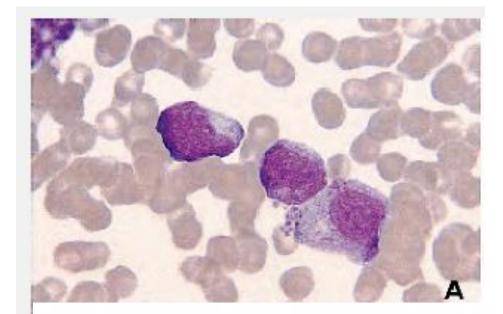
- Nuclei
  - Round
  - Evenly staining
  - 2-3 nucleoli
- Cytoplasm
  - No granules\*
  - Unevenly staining
  - Perinuclear clearing



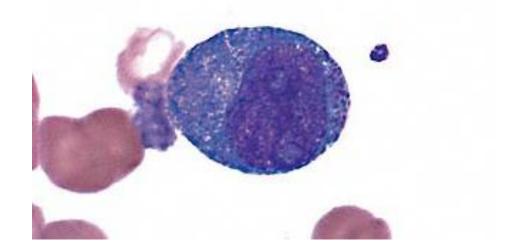


## Myeloblast

• CD34+, CD117+, CD38+, HLA-DR+



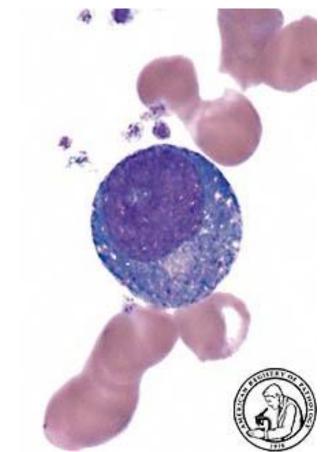
- Also as myeloid cells:
  - CD13+
  - CD33+



## Promyelocyte

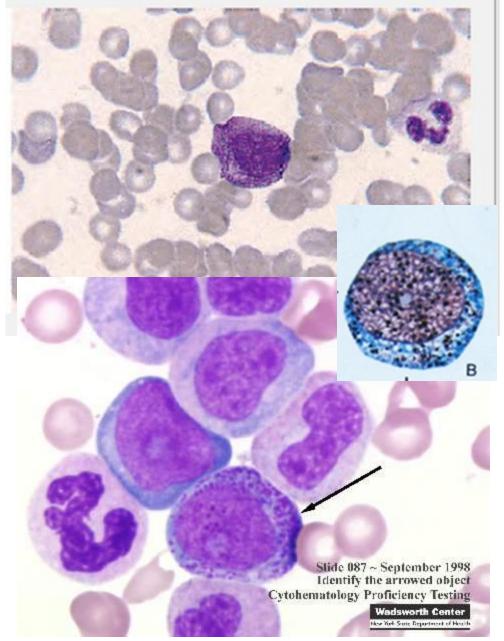
- A myeloblast when develop distinctive granules
- Nucleous
  - Chromatin is coarser than myeloblast
  - Indistinct nucleoli
  - Oval round nuclei
- Cytoplasm
  - Primary granules are darkblue or reddish-blue

Becomes a **myelocyte** once you identify the granules as basophilic, eosinophilic or neutrophilic



## Promyelocyte

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  - Oval round nuclei
- Cytoplasm
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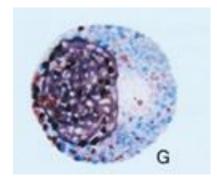
## Neutrophilic Myelocyte

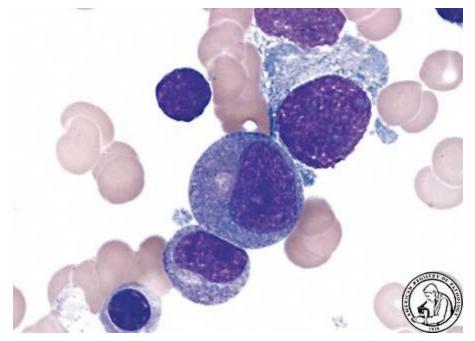
#### Nuclei

 Round to oval, flattened in one side

#### Cytoplasm

 Initially a perinuclear island of ill-defined reddish granules so neutrophilic differentiation

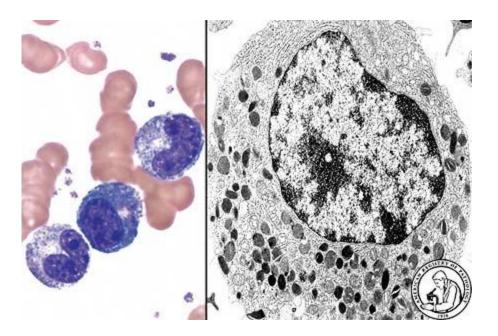




Secondary granule formation begins in the Golgi region highlighted by the paranuclear hof in this early neutrophilic myelocyte (bone marrow aspirate; Wright stain).

## Neutrophilic metamyelocyte

- Nuclei
  - Slightly indented nuclei
- CytoplasmSmall pinkish granules

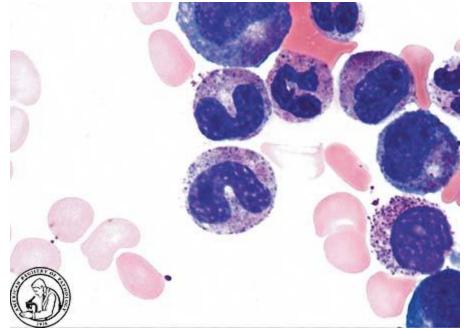


 May be seen normally on PB but also indicates myeloid hyperplasia

Composite of a neutrophilic myelocyte (center), neutrophilic metamyelocyte (top), and band neutrophil (bottom) in a bone marrow aspirate smear shows the progression of maturation changes of the nucleus and cytoplasm. Right: Electron micrograph of a myelocyte shows primary and secondary granules (bone marrow aspirate; Wright stain).

## **Neutrophilic Band**

- Nucleus
  - Characteristic horseshoe nucleus
  - Indentation is greater than half of the hypothetical round nucleus
- Cytoplasm
  - Evenly distributed granules that stain shades of pink and blue
- 1-5% of PB in healthy individuals, if increased is called "shift to the left"

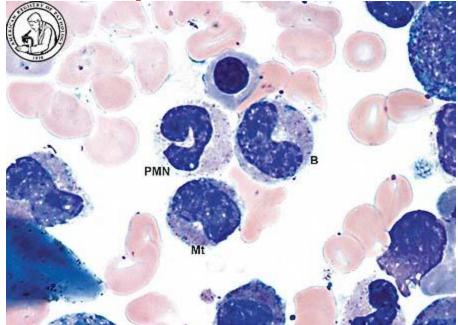


## And finally... The Neutrophil

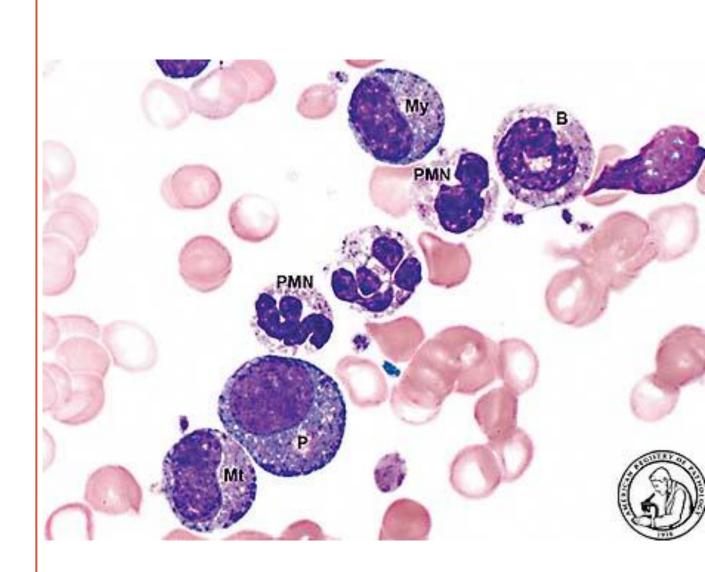
#### Nuclei

- Typically lobulated
- 35% 2 lobes
- 41% 3 lobes
- Hyperlobulation
   →pernicious anemia (6 or more lobes)
- Hypolobulation → Pelger-Huet anomaly (2 round lobes connected with a short filament [pince-nez form])

#### • 50-70% of WBC

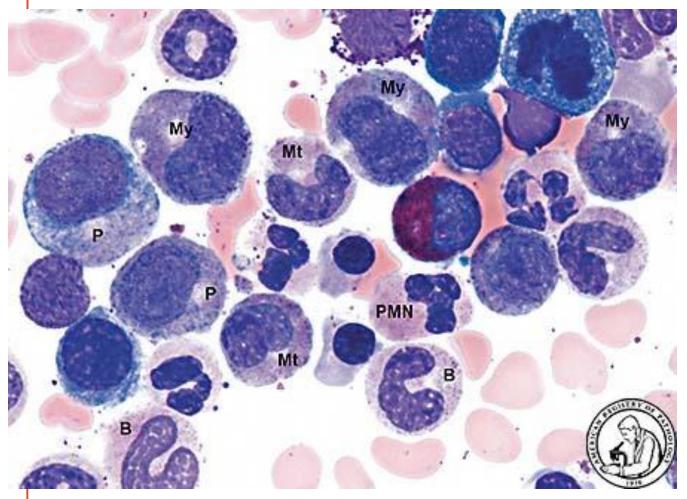


The neutrophilic metamyelocyte (Mt), band neutrophil (B), and segmented neutrophil (PMN) evident in the center of this bone marrow aspirate smear highlight the nuclear and cytoplasmic features of maturation (Wright stain). Progressive nuclear maturation from the round eccentric nucleus of a promyelocyte (P) (lower left) through the myelocyte (My), metamyelocyte (Mt), band (B), and segmented neutrophil (PMN) (bone marrow aspirate smear; Wright stain).



#### STAGES OF NEUTROPHILIC MATURATION

Early promyelocyte (P), neutrophilic myelocyte (My), neutrophilic metamyelocyte (Mt), band neutrophil (B), and segmented neutrophil (PMN). The progression from basophilic to eosinophilic cytoplasm and the acquisition of first, primary, and then, secondary granules in conjunction with gradual and progressive nuclear segmentation and condensation of the nuclear chromatin are evident.

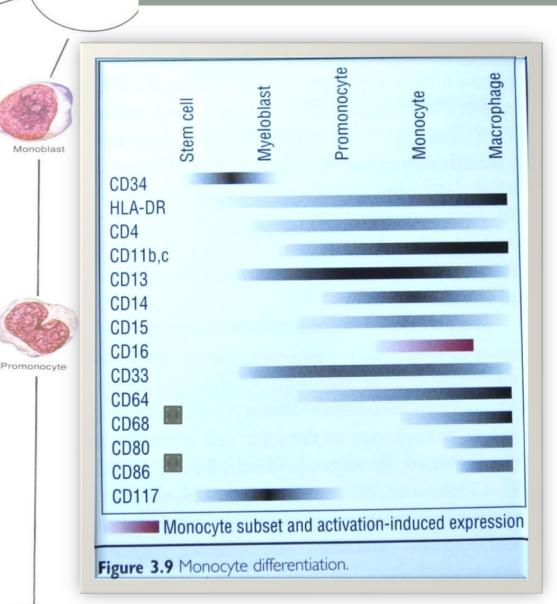


#### Morphological abnormalities

- Toxic granules: prominent blue-black to purplish granules that resemble primary granules. Mean asynchrony between maturation of nuclei and lysosomes
- Dohle bodies: pale skyblue cytoplasmic inclusions (Rough endoplasmic reticulum), acute phase reaction and May-Hegglin anomaly

#### Monocytes

- Monocytes and related dendritic cells play a pivotal role in host defense from microbial pathogens, wound healing, angiogenesis, hematopoiesis, and various inflammatory reactions
- Monocytic production within bone marrow is estimated to take about 2 to 3 days



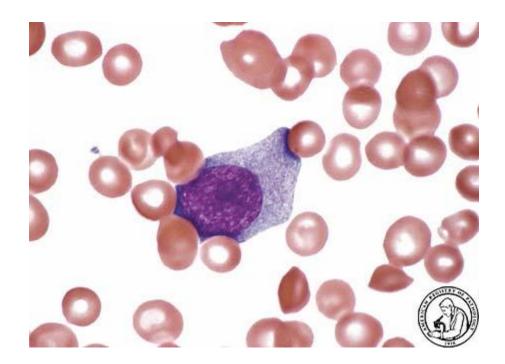


Monocyte

Pluripotential Stem Cell

#### Monocytes

- Neither monoblasts nor promonocytes are typically evident in normal bone marrow
- Monoblasts:
  - Nuclei: round to oval with dispersed, blastic nuclear chromatin
  - Cytoplasm: abundant and pale blue, with either agranular or subtle, finely granular cytoplasm

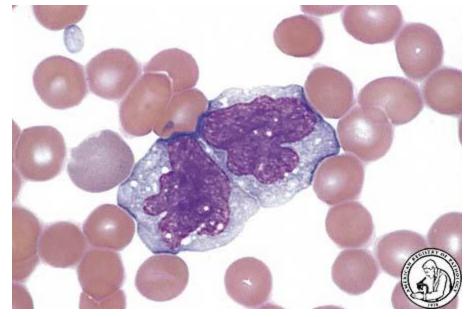


Leukemic monoblast has voluminous, slate blue-gray, finely granular cytoplasm and an immature round nucleus

### Monocytes

#### Promonocytes:

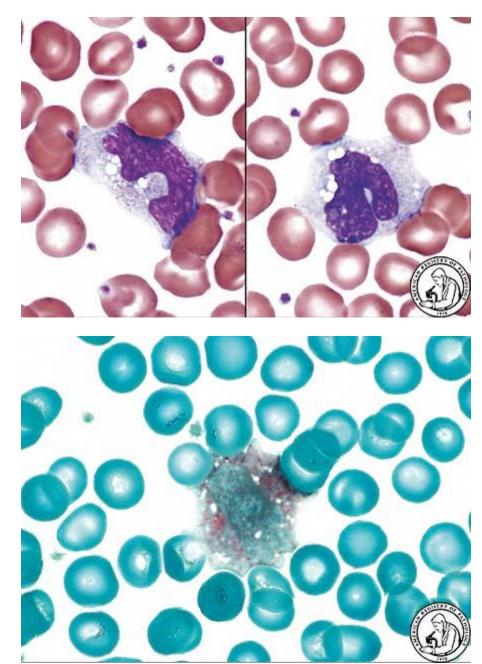
- Nuclei: folded nuclear configuration with a typically prominent nucleolus and fairly dispersed nuclear chromatin
- Cytoplasm: abundant and pale blue, with either agranular or subtle, finely granular cytoplasm



Leukemic promonocytes have abundant cytoplasm and folded, immature nuclei

#### Monocytes/Macrophages

- Diffuse cytoplasmic positivity for alphanaphthyl butyrate and alpha-naphthyl acetate esterase (non-specific esterases) in all monocyte stages
- IHC include:
  - Lysozyme, CD68 (KP1 or PG-M1 epitopes), and CD163



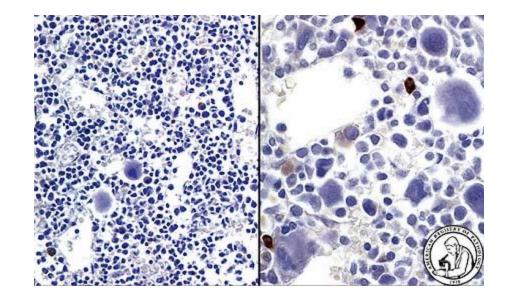
#### Dendritic cells

 Dendritic cells are defined more by their functional activities than by specific morphologic features, although immunophenotypic subsets are well described

 Dendritic cells varies by the specific cell type, and differences in phenotype are based on derivation from either myeloid or lymphoid progenitor cells

#### Dendritic cells

 Dendritic cells are infrequent in bone marrow, and immunohistochemic al techniques (a profile consisting of CD68, CD123-, and CD43-positive, myeloperoxidasenegative cells) are generally required for cell identification

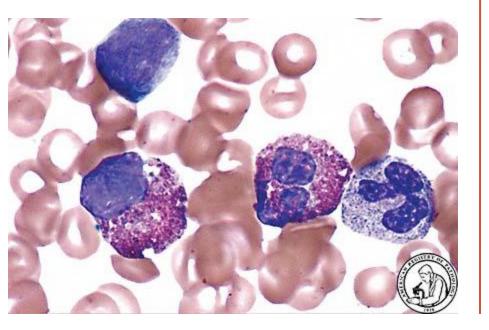


 This composite shows rare CD1apositive cells (left) and slightly more numerous S-100 protein-positive cells (right) in a normal bone marrow core biopsy from 1-year-old female

# Other myeloid components

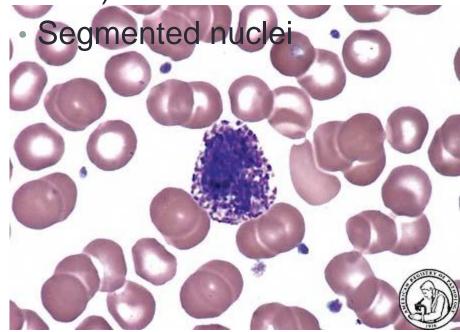
### Eosinophilic myelocyte and mature eosinophil

 Eosinophil granules are large and refractile, and contain major basic protein, eosinophil peroxidase



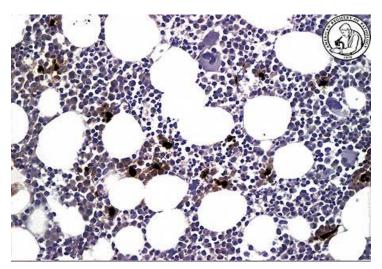
#### Mature segmented basophil

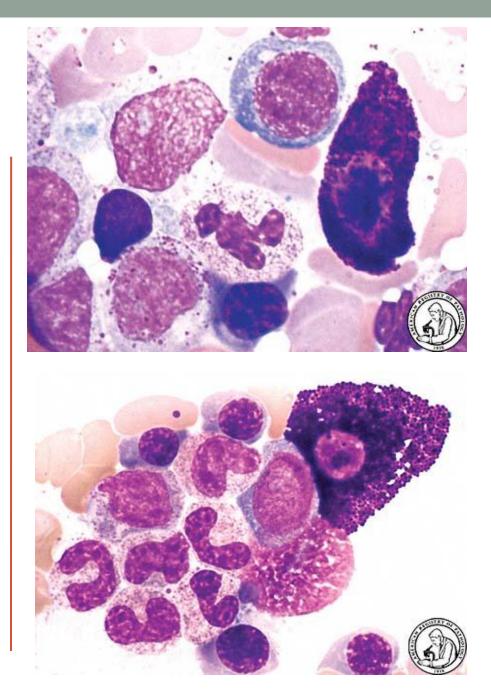
 Granules have histamine and heparin (like mast cells)

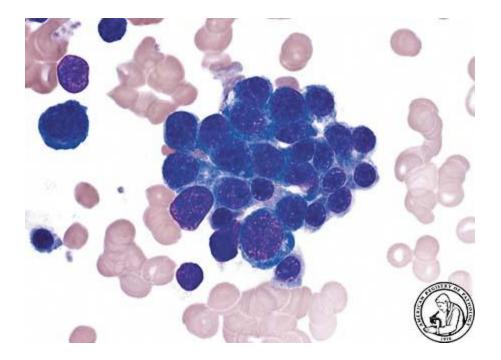


# Mast Cells

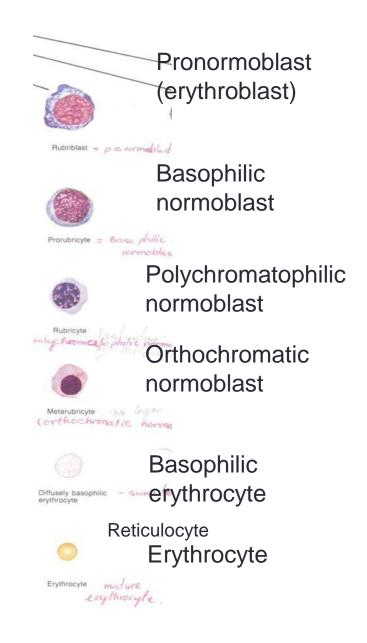
- Round to oval nuclei
- Granules have histamine and heparin
- Tryptase or CD117, highlight perivascular distribution in bone marrow



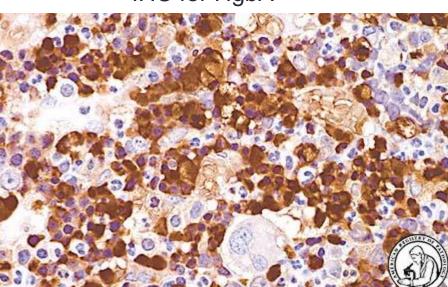


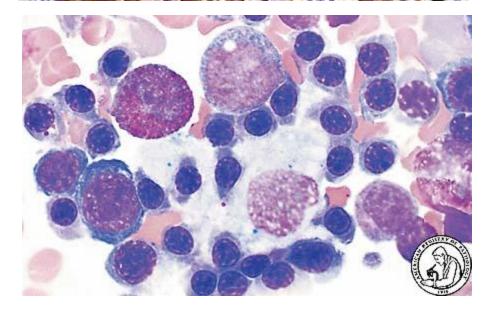


Erythropoiesis occurs in discrete colonies within the hematopoietic cavity Basophilic RNA-rich cytoplasm in immature forms Production takes 5 to 7 days



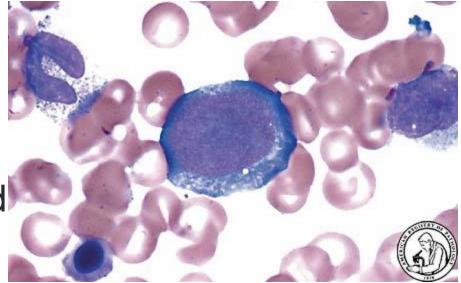
- Erythropoietin is the dominant lineage specific growth factor
- Controls survival and proliferation
- Binds to a receptor that activates JAK2, initiating the downstream signaling pathways resulting in phosphorylation and activation of GATA1

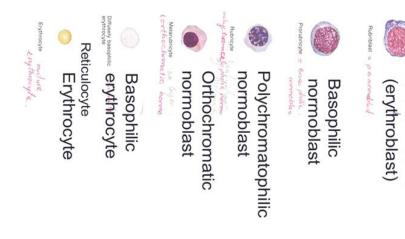


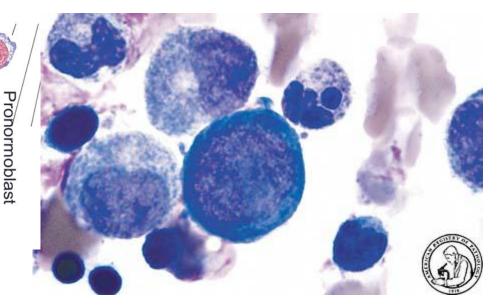


IHC for HgbA

 erythroblast has deeply basophilic cytoplasm and a round nuclear contour



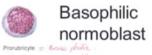




Basophilic normoblasts

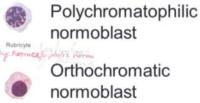




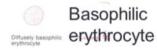




Rubricyte

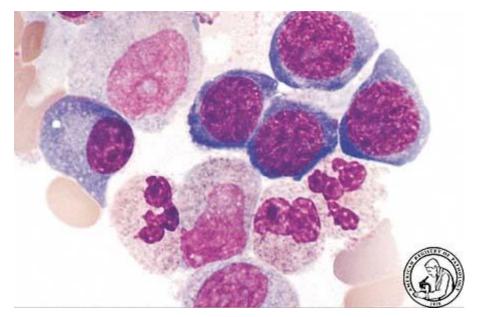


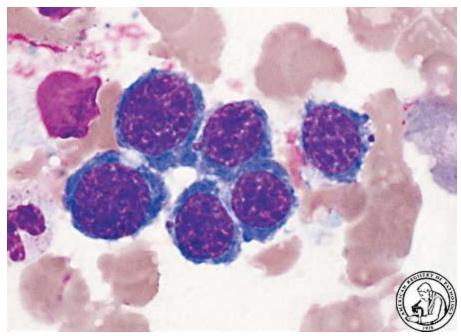
Metarubricyte the Legar (orthochromatic norma



Reticulocyte Erythrocyte

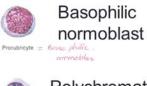
Erythrocyte mature empthrocyte.

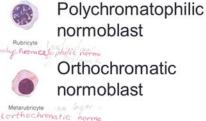




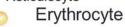
- Polychromatophilic normoblasts
  - Last stage of cellular division





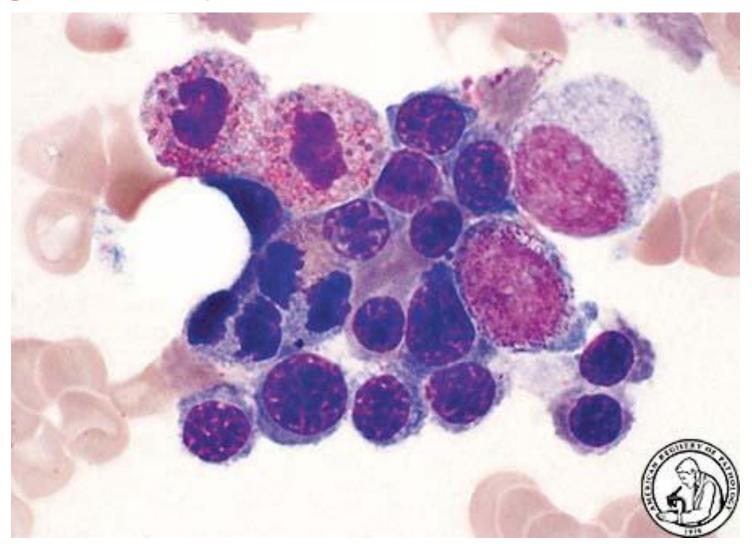






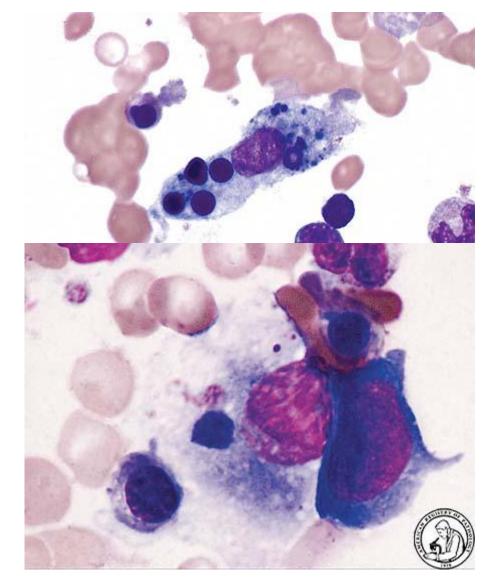
Erythrocyte mature enythrocyte

#### **Stages of Erythroid Maturation**



#### Monocytes/macrophages





- Megakaryoblast
- Promegakaryocyte
- Platelet-shedding megakaryocyte
- IHC:
- CD61, CD41, CD42b





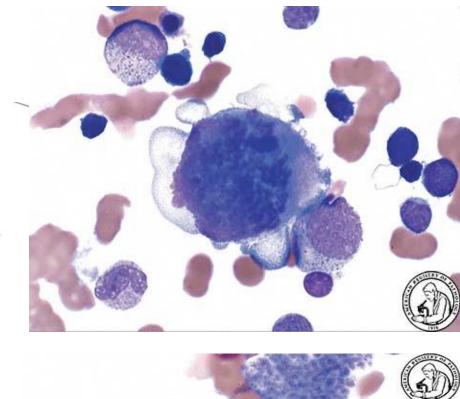








Thrombocytes



# Megakaryocyte Morphology

Stage of Maturation	Morphology	
Megakaryoblast	Difficult to recognize by morphology alone, but tends to be large blast with a high	
	nuclear to cytoplasmic ratio, basophilic	Megakaryoblast
	cytoplasm, and variable cytoplasmic blebbing	
Promegakaryocyte	Spectrum of large cells with various degrees	Promegakaryocyte
	of nuclear lobulation	
	Progressive increase in overall size, variable	200
	cytoplasmic granules	Megakaryocyte without thrombocytes
Platelet-shedding	Large multilobulated megakaryocytes with	minout shortbodyas
megakaryocyte	highly condensed nuclear chromatin	
	reside adjacent to bone marrow sinuses	
	Voluminous amounts of cytoplasm with	
	abundant cytoplasmic granules	Metamegakaryocyte
		100

Thrombocytes



Megakaryoblast





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Megakaryocyte without thrombocytes

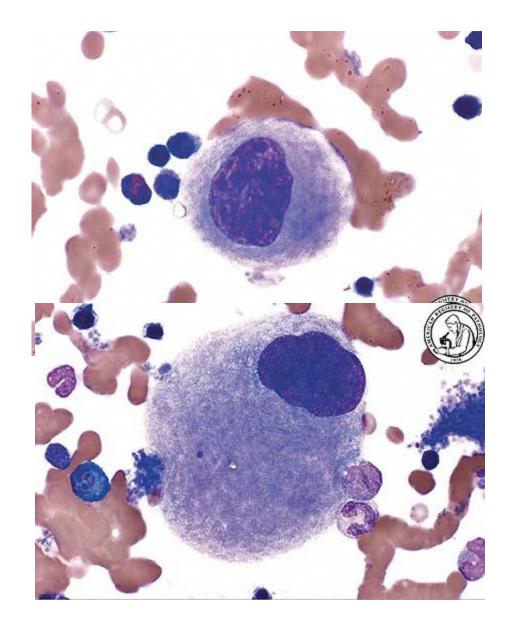
Stage of Maturation	Cytochemical/Immunophenotypic Properties	
Megakaryoblast	Platelet peroxidase evident by immuno-electron microscopic techniques Variable CD34 expression	
	Expression of lineage-specific antigens such as CD41, CD42b, CD61, and Mp11	
Promegakaryocyte	Loss of CD34 but retention of the full complement of megakaryocyte- associated antigens	



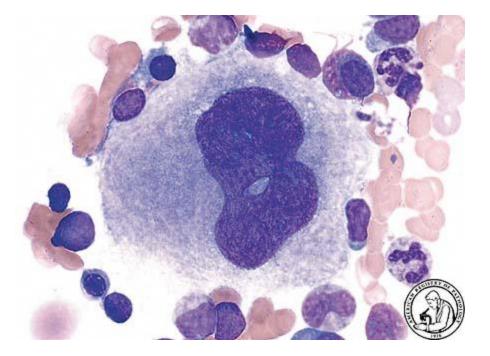
Thrombocytes

Platelet-shedding megakaryocyte Expression of some megakaryocyte-associated antigens such as CD31, CD41, vWF increases with maturation

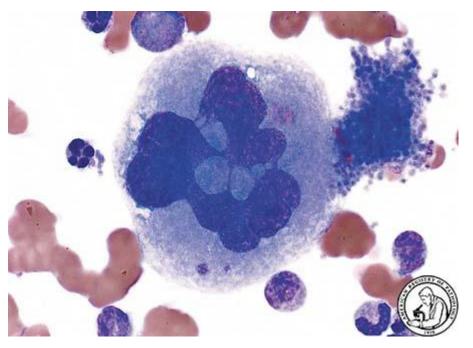
 Early to middle maturation stage megakaryocyte with round nucleus and abundant cytoplasm



 Megakaryocyte maturation is characterized by the progressive doubling of nuclear material, with the multilobulation of a single nucleus, termed endomitosis



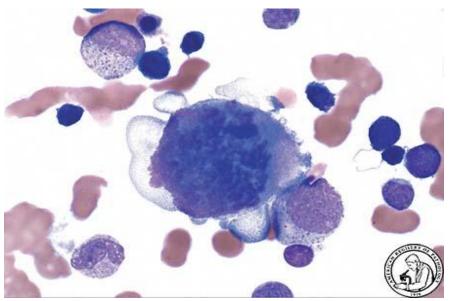
Prominent nuclear
 lobulation with
 interconnected lobules
 is evident in this
 mature megakaryocyte
 with an adherent large
 platelet clump



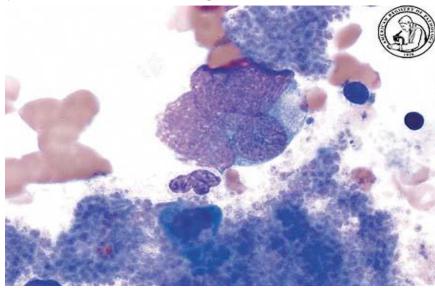
 Thrombopoietin (produced in the liver) is an obligatory lineage specific growth factor

 Binding of thrombopoietin to its ligand (c-Mpl) activates JAK2 → promotes meg differentiation and proliferation

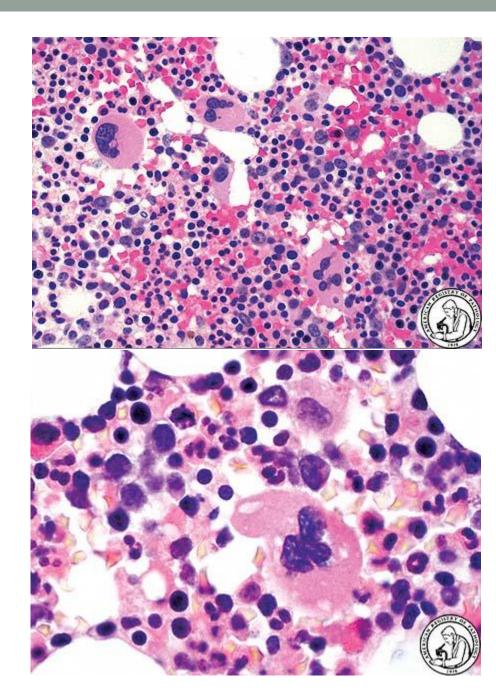
 Platelets have thrombopoietin receptors



Immature megakaryocytes with prominent cytoplasmic blebbing

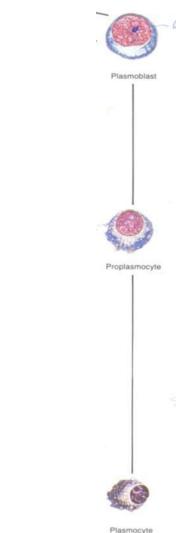


 Perisinusoidal and intrasinusoidal localization of megakaryocytes is evident in this bone marrow biopsy from an adult



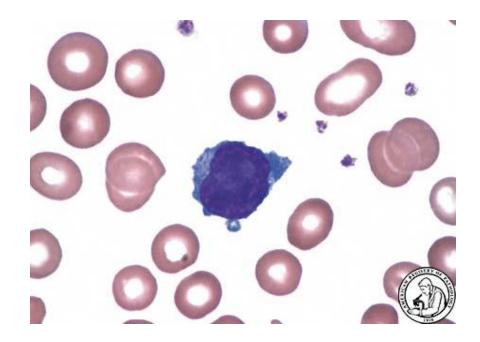
# Lymphoid and Natural Killer Cells

- Origin of B, T, and natural killer (NK) precursor cells from stem cells that give rise to hematopoietic and lymphoid lineages
- Immature T cells migrate to the thymus
- The bone marrow is the site of B-cell development



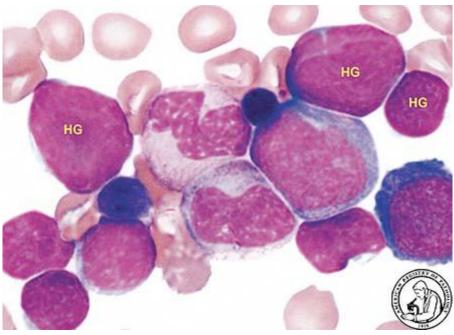
#### Hematogones

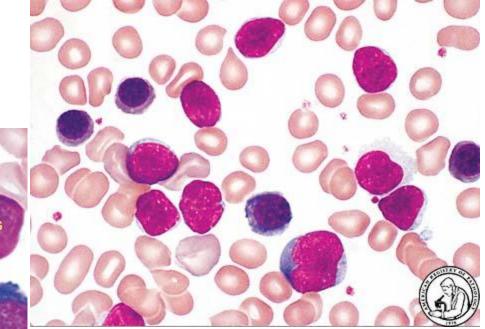
(immature benign B lymphoid cells), may be abundant in specimens from pediatric patients



 An immature lymphoid cell with dispersed chromatin and an irregular nuclear configuration is evident in the peripheral blood of a normal infant

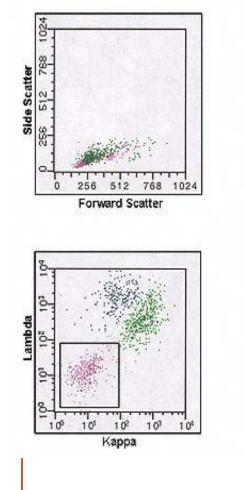
- BM Hematogones
  - Nuclear chromatin is highly condensed

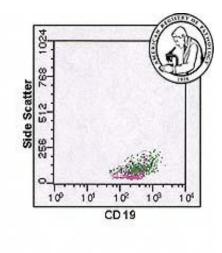


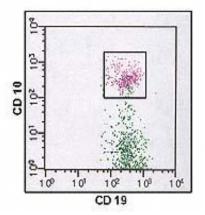


#### Hematogones

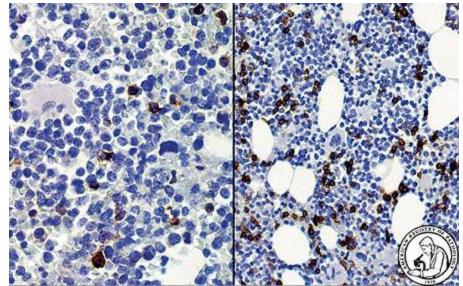
 By gating the lymphocyte population by forward and side scatter properties, an admixture of mature polyclonal B cells and surface immunoglobulinnegative, CD10-positive hematogones is evident in a normal bone marrow aspirate





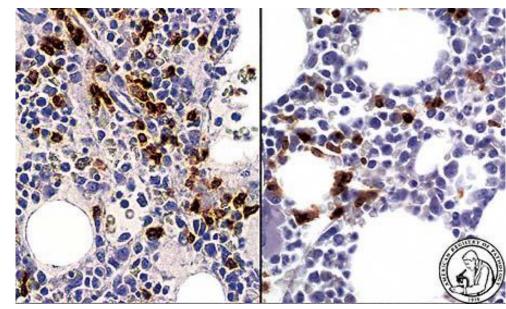


 bone marrow clot section from a premature infant with increased hematogones shows a side-by-side comparison of the number of CD3positive T cells (left) and the number of CD20positive B cells (right). Because of the increased numbers of hematogones, B cells are more numerous than T cells



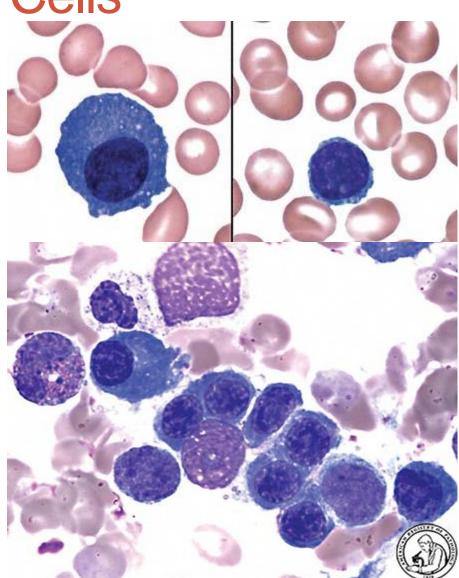
 Except in specimens with abundant hematogones, T cells predominate in normal bone marrow and exhibit a patchy, partially perivascular distribution

 B cells are less common and are randomly distributed and dispersed, individually or in small clusters

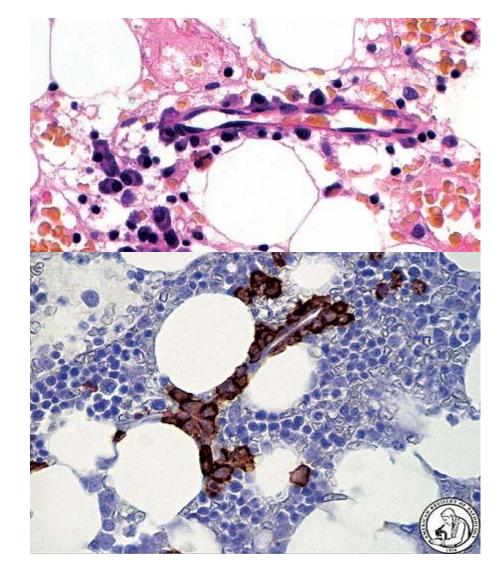


 The perivascular (left) and patchy interstitial (right) distribution of T cells is evident on the bone marrow core biopsy from an adult.

- Mature plasma cells
  - Nuclei: eccentric with a "clockface" chromatin pattern
  - Cytoplasm: prominent paranuclear hof, and abundant basophilic cytoplasm that may contain immunoglobulin vacuoles

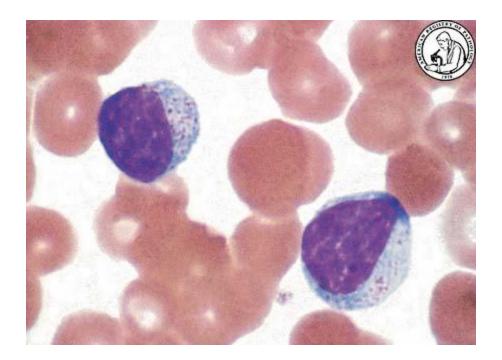


- Plasma cells may have perivascular distribution
- CD138 highlights perivascular plasma cells
- Constitute up to 5% of plasma cells
- Benign plasma cells have a polytypic kappa to lambda light chain ratio



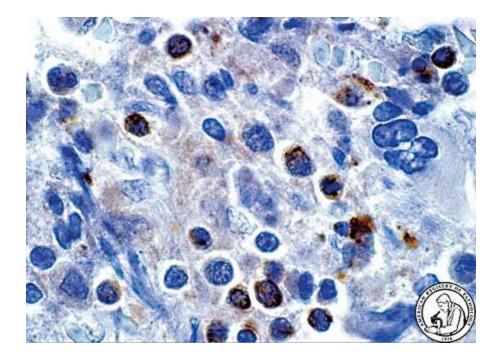
### NK Cells

- NK cells are surface CD3-negative, CD56positive, CD16-positive
- Produce immunoregulatory cytokines
- Also mediate cytotoxicity against target cells that lack matching major histocompatibility complex (MHC) ligands



- NK cells have features of large granular lymphocytes, although cytotoxic T cells (surface CD3 and CD8 positive) also share this morphology
- The granules contain perforin and granzymes,
- IHC: granzyme and T-cell intracellular antigen 1 (TIA-1)

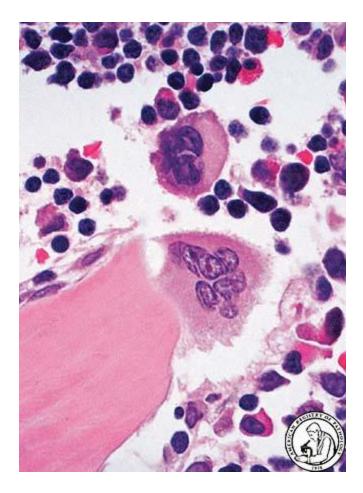
 TIA-1 IHC stain highlights cytoplasmic granules in a bone marrow core biopsy with increased large granular lymphocytes.



#### **Bone Elements**

3 elements:

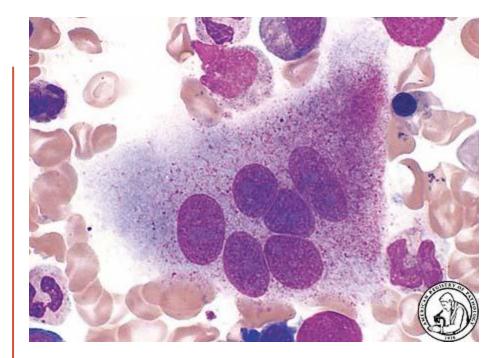
- Osteoblasts
- Osteoclasts
- Osteocytes
  - Osteoblasts assemble to form new bone in the lacunar spaces vacated by osteoclasts, a process called coupling



#### **Bone Elements**

Osteoclast

- Osteoclasts resorb bone and are derived from a common monocytic/ macrophage/ dendritic progenitor cell
- Monocytic cells differentiate into osteoclasts in the presence of M-CSF and RANKL

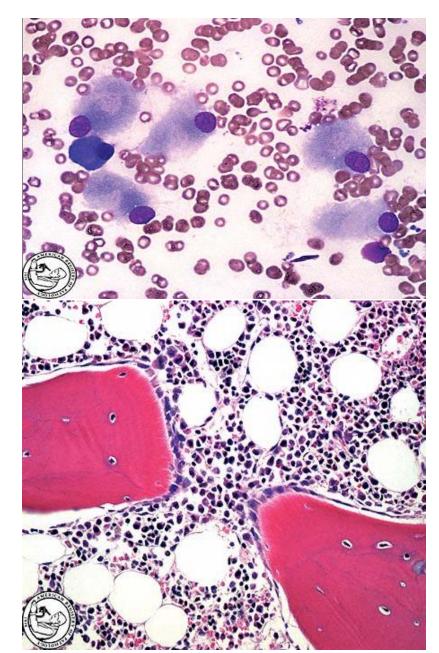


 Discrete nuclei, coarse cytoplasmic "bone sand" in mature cells, and paratrabecular localization often within scalloped spaces

### **Bone Elements**

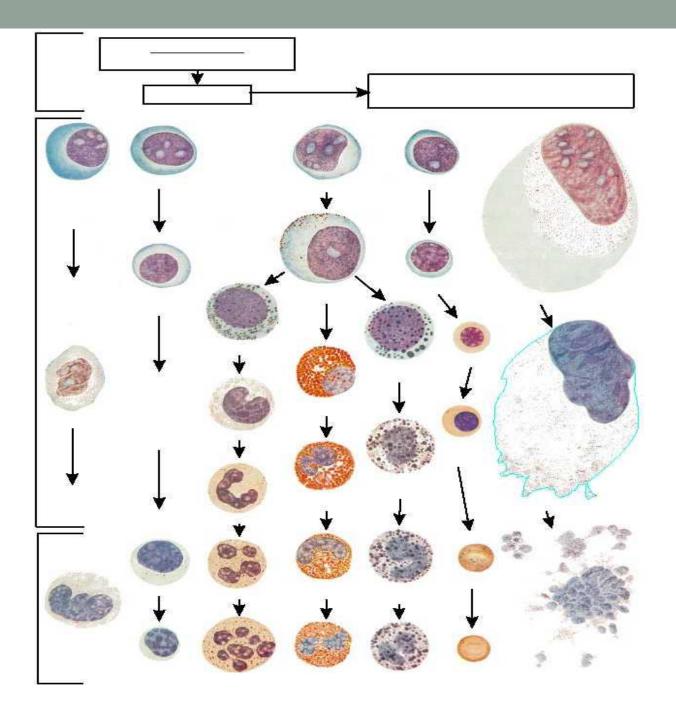
#### Osteoblast

- Stromal-derived cells which produce the bony substrate
- Resemble enlarged plasma cells
- Osteoblasts also contain a cytoplasmic pale area (hof) which, unlike plasma cells, is separated from the nucleus
- If active bone remodeling is in progress, osteoblasts rim bony trabeculae in a single file



#### Age Related Changes

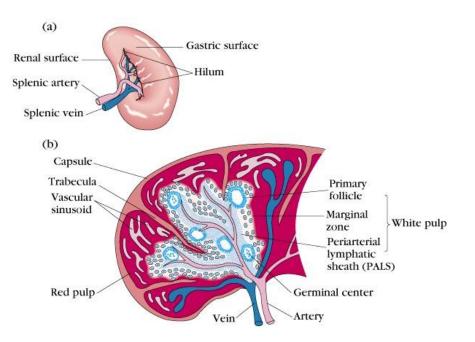
Patient Age	Cellularity	Bony Trabeculae	Cellular Composition
Newborn	Up to 100%, but may be lower	Very active bone remodeling and incomplete ossification of cortical bone	Blasts up to 5% Predominance of myeloid cells except in cases in which hematogones are numerous; myeloid to erythroid ratio of ~ 4:1 Lymphoid cells, notably hematogones, may be numerous
Infant	Variable, up to 100%, but may be lower	Very active bone remodeling and incomplete ossification of cortical bone	Blasts up to 5% Predominance of myeloid elements; myeloid to erythroid ratio: ~ 5-10:1 Erythroid elements markedly reduced during physiologic nadir Lymphoid cells, notably hematogones, may be abundant (up to 50% of cells)
Child	60-80%	Active bone remodeling	Blasts up to 5 percent, but usually lower Myeloid elements predominate; myeloid to erythroid ratio: ~ 3:1 Lymphocytes, notably hematogones, may be abundant
Young adult	50-70%	Bone remodeling may be evident, especially in young males	Blasts generally <5% Myeloid elements predominate; myeloid to erythroid ratio: ~ 3:1 Lymphocytes generally inconspicuous, but may range up to 20%
Adult	40-60%	Bone remodeling absent Osteoclasts and osteoblasts inconspicuous Bony trabeculae may be thinned (osteopenic), especially in females	Blasts generally <3% Myeloid elements predominate; myeloid to erythroid ratio: ~ 3-4:1 Lymphocytes usually inconspicuous, but may range up to 20%
Elderly	25-40%	Bone remodeling absent Osteoblasts and osteoclasts inconspicuous Bony trabeculae may be thinned (osteopenic), especially in females	Blasts <3% Myeloid elements predominate; myeloid to erythroid ratio: ~ 3-4:1 Mild dysplastic features may be noted Lymphocytes are generally inconspicuous, but may range up to 20%, especially if hematopoiesis is reduced. Lipogranulomas and lymphoid aggregates may be present



# HISTOLOGY OF THE SPLEEN

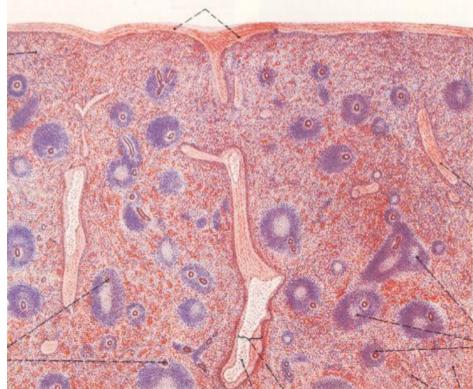
# **Gross Anatomy**

- Normal weight 150 g
  - SD 25 g
- Hilus, where it is penetrated by vessels and nerves which follow the extensive branching network of fibrous trabeculae.
- Accessory spleens occur in about 10 percent of individuals
- Following traumatic rupture, small nodules of splenic tissue may grow on the peritoneal surface as implants (splenosis)



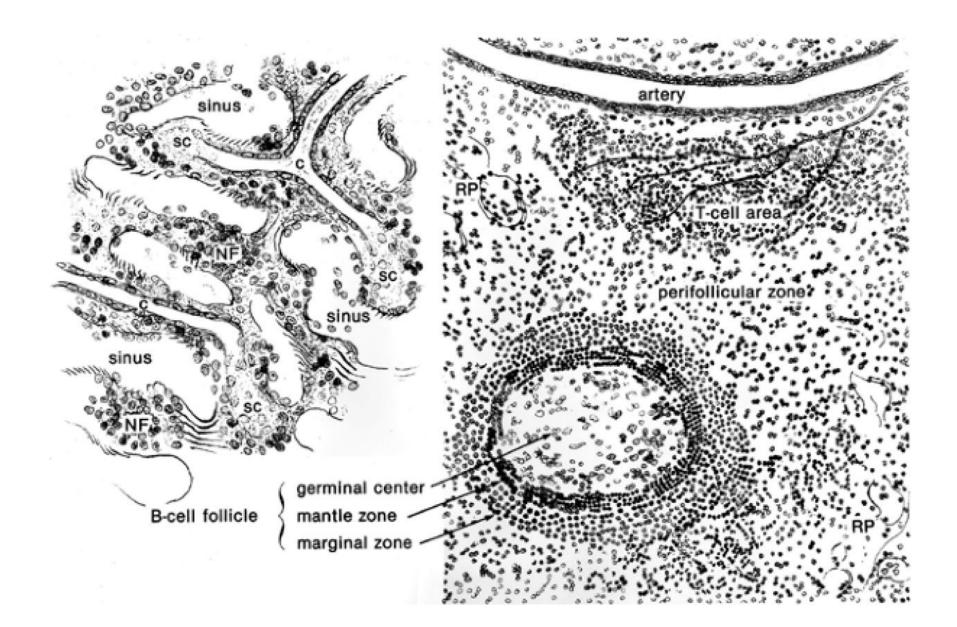
# White Pulp

 Comprises the lymphoid compartment of the spleen and consists of both
 follicular B-cell-rich areas as well as Tcell-rich periarteriolar
 lymphoid sheaths.



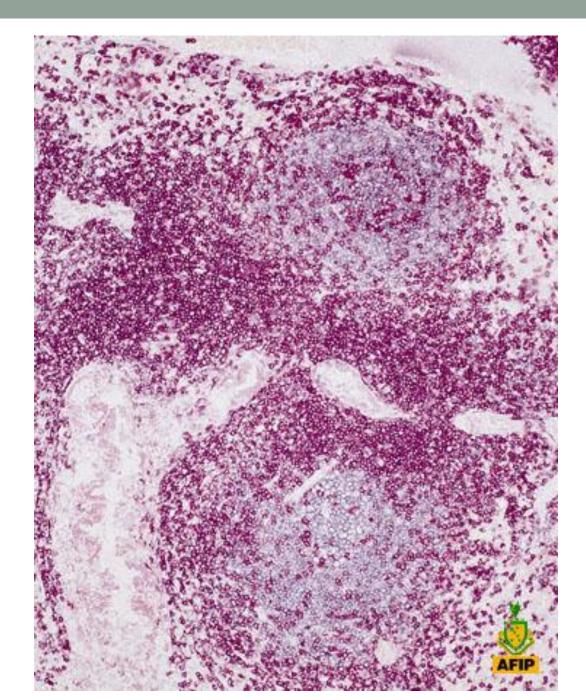
### Periarteriolar T-Cell-Rich Lymphoid Sheaths

- Counterpart to the paracortical region of lymph nodes
- Lymphoid sheath, which surrounds splenic arteries as they exit the fibrous trabeculae



#### Spleen: Periarteriolar area

The T cells predominate in the periarteriolar lymphoid sheath (labeled red with Leu-22/ CD43). The follicles, which tend to occur at arterial branch points, are labeled blue (L26/CD20).



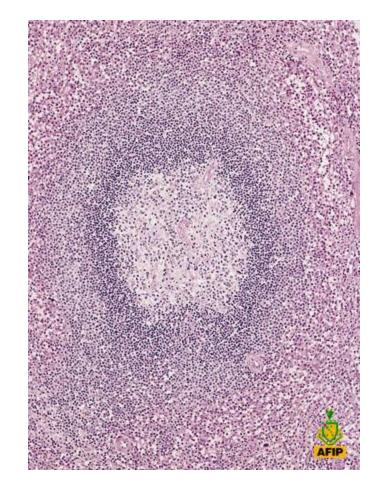
### **Primary and Secondary B-Cell Follicles**

 Located at the periphery of the T zone and have the identical histologic and phenotypic features of primary and secondary follicles of lymph nodes

### **Primary and Secondary B-Cell Follicles**

#### MARGINAL ZONE

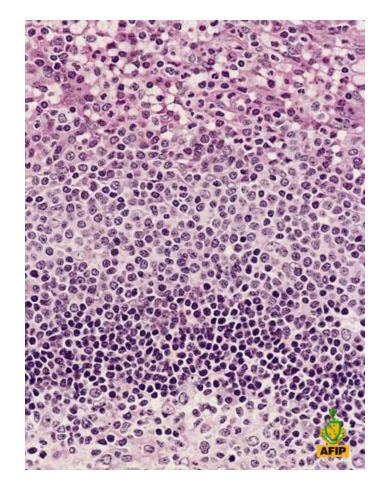
- Surrounds the primary follicle and the mantle zone of secondary follicles
- Consists of a corona of medium-sized lymphoid cells with prominent pale cytoplasm



### **Primary and Secondary B-Cell Follicles**

#### MARGINAL ZONE

- The nuclear chromatin of the intermediate-sized marginal zone cells is somewhat less condensed than that of small lymphocytes
- Admixed with a variable number of plasma cells, T cells, and macrophages



### Red Pulp

- 4 vascular structures:
  - Slender and nonanastomosing arterial vessels
  - Reticular meshwork of thin plates of cellular tissue lying between the sinusoids comprising splenic cords
  - Large, thin-walled venous vessels called sinusoids

.

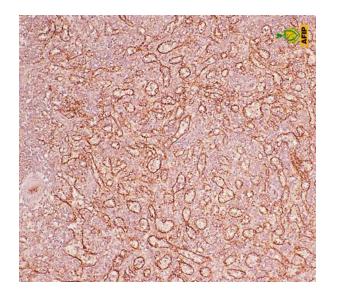
 Pulp veins which drain the sinusoids

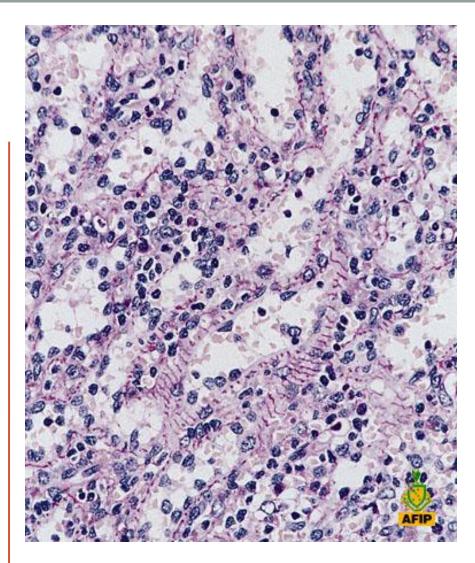
### Cords

- Reticular meshwork consists of a branching system of cords lying between the sinuses
- Includes the reticular meshwork and may run through a sheath of macrophages with may run through a sheath of macrophages
- Clearance functions are also handled by marginal zone and red pulp macrophages

### Sinusoids

- Are lined by tapered endothelial cells separated by slit-like spaces and surrounded by distinctive ring fibers and bridging fibers
- Stain endothelial markers (FVIII) and CD8

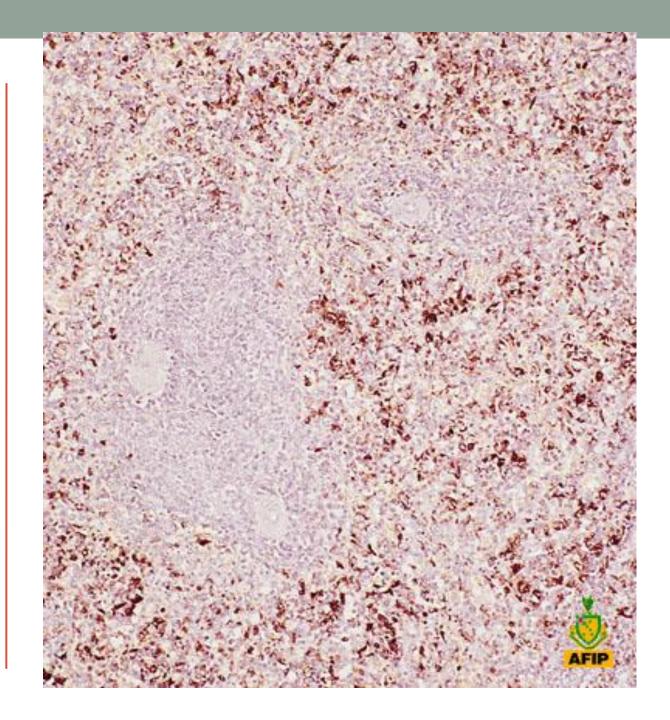




 PAS stain highlights the distinctive ring fibers and bridging fibers

#### Splenic macrophages

Macrophages are preferentially located in the marginal zone and red pulp cords of the spleen (labeled brown with KP-1/CD68).



# Spleen, physiology

- Differentiation of reticulocytes, platelets, and monocytes
- Removal of abnormal erythrocytes
- Major site of antibody production, particularly in response to bloodborne antigens.

# **Useful References**

#### Leukemias

- WHO 2008
- For a single author perspective:
  - Bone marrow -> Dr. Foucar or Dr. Bain books
  - Online free AFIP book:
  - <u>https://www1.askafip.org/port</u> <u>al/page?\_pageid=33,319226</u> <u>&\_dad=portal&\_schema=PO</u> <u>RTAL&pAction=PREVIEW\_P</u> <u>AGE&pBook=3F14&pPage=</u> <u>8</u>

#### Lymphomas

- WHO 2008
- For an algorithmic approach → <u>http://www.uscap.org/newi</u> <u>ndex.htm?98th/education.</u> <u>htm</u>
- For benign lymph node
   →loachim's Lymph node pathology

### Based on...

 AFIP Atlas of non-tumor pathology – Bone marrow by Dr. Foucar.